

Proteomic platform for comprehensive and quantitative urinary proteomes

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Urine is a desirable body fluid for clinical research as it can be obtained non-invasively in large quantities from every patient. Proteins in urine are mainly derived from plasma or are secreted from kidney and the urinary tract, providing a potential read-out for various diseases and representing an attractive source for biomarkers.

Mass spectrometry (MS)-based proteomics could in principle be an ideal technology to discover and detect protein biomarkers in this clinically accessible and relevant body fluid. However, analysis of urine is a challenging task due to the high levels of salt and other interfering compounds, low protein concentrations and a high degree of intra- and inter-individual variability.

Here we build on recent developments in our group in the sample-preparation workflow¹ to enable reproducible, parallelized and sensitive processing of urine proteins. We collected urine samples from 6 healthy male donors on 3 consecutive days. We identified a total of 3284 proteins in urine and calculated the label-free quantification values² of 2200 proteins in average per sample, with MS-signals spanning 6 orders of magnitude. Remarkably, we identified 1354 proteins, which have not been reported in urine before and these are enriched for terms such as “humoral immune response”. Encouragingly, a common set of 1112 urine proteins (‘core proteome’) were detectable in all studied individuals. Median coefficient of variation (CV) was 27% for technical replicates, which is excellent for label-free shotgun proteomics. The core proteome had a CV of 42% between the 6 individuals.

Our developments contribute to a robust and sensitive high-throughput urine proteomics platform which we hope will open urine proteomics to routine, quantitative analysis of patient samples in clinical settings.

- 1) Kulak NA, Pichler G, Paron I, Nagaraj N, Mann M. Minimal, encapsulated proteomic-sample processing applied to copy-number estimation in eukaryotic cells. *Nat Methods*. 2014 Mar;11(3):319-24. doi: 10.1038/nmeth.2834. Epub 2014 Feb 2.
- 2) Cox J, Hein MY, Luber CA, Paron I, Nagaraj N, Mann M (2014) Accurate proteome-wide label-free quantification by delayed normalization and maximal peptide ratio extraction, termed MaxLFQ. *Mol Cell Proteomics* **13**: 2513-2526