Annika Andersen (1),Julia Remnestål (1) ,Fredrik Edfors (1), Mathias Uhlén (1), Jochen Schwenk (1), Anna Häggmark-Månberg (1), Peter Nilsson (1) and Claudia Fredolini (1)(2)
(1) SciLifeLab, KTH - Royal Institute of Technology, Stockholm, Sweden , (2) Department of Immunology, Genetics and Pathology and SciLifeLab, Uppsala University

## Introduction

Alzheimer's disease is an irreversible, progressive brain disorder and the most common cause of dementia among older adults. Damages in the brain are expected to starts a decade or more before cognitive problems become evident. The identification of biomarkers associated to early biological events in the onset and progression of Alzheimer's disease would be of tremendous importance in both prevention and treatment. Suspension bead arrays (SBA) has proven to be a convenient approach to profile hundreds of proteins in large cohorts of biological samples. The aim of this study was to develop and validate Parallel Reaction Monitoring (PRM) assays to perform an orthogonal verification of clinically rlevant protein profiles previously discoved by SBA[1]
[1] Julia Remnestål et al. Proteomics Clin Appl. 2016 Dec; 10(12): 1242-125

## Material \& Methods



## Figure 1. Graphical representation of the workflow

94 CSF samples were collected at Sahlgrenska Hospital in Gothenburg (Sweden) from patients with AD, preclinical AD (down A $\beta 42$ ), prodromal AD (down AB42, up t-tau or $p$-tau), non-AD dementia, MCl (mild cognitive impairment) and age-matched healthy controls (Table 1). Protein profiling was performed with SBA as previously described by Remnestål et al. Twenthy proteins with interesting profiles were selected for further verification with PRM-MS. CSF samples were spiked with QPrEST internal standards and digested (Trypsin/Lys-C) overnight. Clean-up was performed with solid phase extraction (SPE) containing hydrophilic-lipophilic balance (HLB) sorbent in a $\mu$ Elution plate format (Waters). Liquid chromatography (LC) was used for peptide separation with a 35 -min linear gradient on a 25 cm C18 analytical column (Thermofisher). PRM analysis was performed on a Q-Exactive HF mass spectrometer (ThermoFisher) with a full scan event (mass range $\mathrm{m} / \mathrm{z} 150-2000$, resolution 60000 , AGC value $2 e 5$ and maximum IT 55 ms ) followed by up to 25 PRM events (resolution 30000 , AGC value 1e6, maximum IT 250 ms , precursor isolation window 2.0 Th ). A minimum of three transitions was required for identification and peaks were manually inspected. Light-to-heavy ratio were exported from Skyline and analyzed using the statistical environment R .

## Table 1. Sample demographic

Results






Figure 2. (Left) Chromatograms of PRM transitions.(Right)I Linear regression and 95\% confidence band.

Table 2. Method validation


Figure 3. Box-and-whisker plot visualization of protein profiles.Pairwise comparison between group levels was performed by Wilcoxon Rank um Test with Bonferroni correction.

| GAP43 | 0.78 | 0.76 | 0.56 |
| :--- | :--- | :--- | :--- | :--- | :--- |
| An | 0.97 | 0.81 |  |



VCAM1



Figure 4. Six examples of scatterplots with value of correlation (Pearson's coefficient) of protein levels measured by suspension bead arrays (MFI) and PRM (L/H ratio).

## Conclusions

- PRM assays for the quantification of 13 brain enriched proteins in CSF were developed and validated. - Quantification by PRM confirmed the profiles observed by SBA for 7 proteins: GAP43, SERPINA3, LRG, APOA4 NEFM, VCAM and CCK
-PRM dad strongy suggest GAP43,VCAM and CCK as CSF molecular markers of early onset of Alzheimer's dise - Peptide quantification in digested crude CSF show a sensitivity in the range of high $\mathrm{ng} / \mathrm{mL} \mathrm{to} \mathrm{mg} / \mathrm{mL}$. Therefore,further method development is required to obtain an accurate quantificaton of low abundant proteins such as CCK, eventually including immuno-enrichment.

