Analysis of the site-specific N-glycosylation of the HeLa cell lysate

<u>Simon Sugár</u>, András Ács, Ágnes Gömöry, Gábor Tóth, Károly Vékey, László Drahos, Lilla Turiák

MS Proteomics Research Group, Hungarian Academy of Sciences, Research Centre for Natural Sciences, Budapest, Hungary

Objectives:

- I. Analysis of the site-specific *N*-glycosylation of HeLa proteins
- II. Assessment of the degree of contamination originating from the culture media

Introduction:

• HeLa cell lines:

derived from cervical cancer tissue, commonly used in cancer research and as an MS standard, commercially available in lysate form

Conclusions:

- I. First in-depth study of site-specific HeLa *N*-glycosylation – enhanced reliability of results and novel information on HeLa *N*-glycosylation
- II. High degree of contamination from culture media causes significant bias – HeLa glycosylation should only be studied in a site-specific manner

Bovine contamination:

Commercially available HeLa cell digest (Thermo Scientific, Waltham, USA) contained high amounts of bovine protein contaminants likely from the FBS culture media

HeLa N-glycosylation:

information on the *N*-glycosylation of HeLa proteins is lacking altough glycosylation carries potentially important biological information

 Site-specific glycopeptide analysis: glycopeptides are analyzed in intact form opposed to analysing the glycan and peptide moieties separately – more valuable information but more challenging due to lower abundance as opposed to non-glycosylated peptides

Workflow:

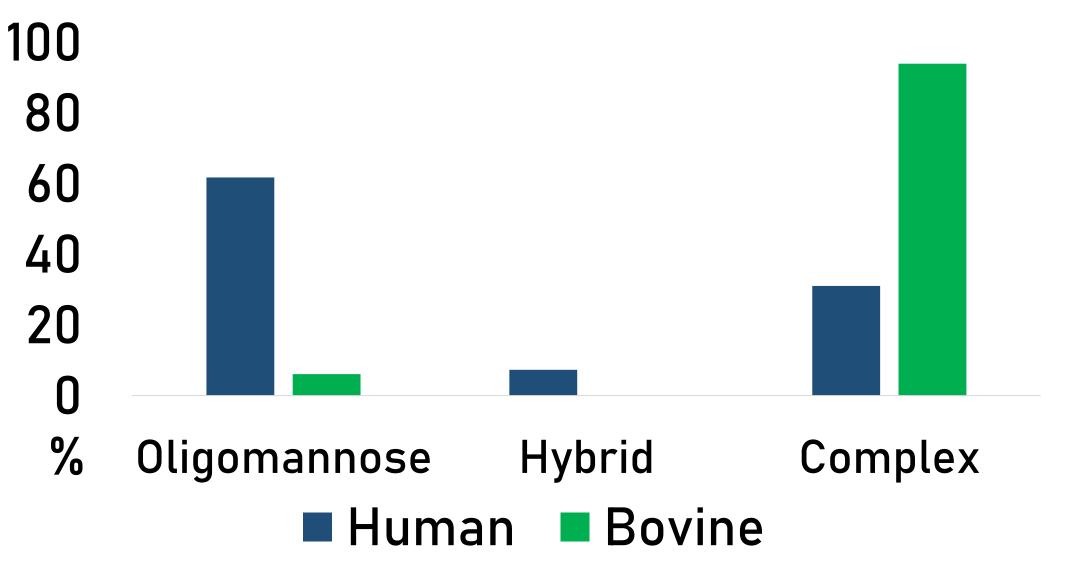
- Complex workflow: glycopeptide enrichment followed by identification of components via HPLC-DDA-MS/MS and label free quantitation using HPLC-MS data using our in-house built GlycoPattern software
- The focus was not on maximizing the quantity of identified glycopeptides but on enchanced reliability
- Introduction of a multi-step manual data validation process, which involved the evaluation of specific high and low energy MS/MS spectra

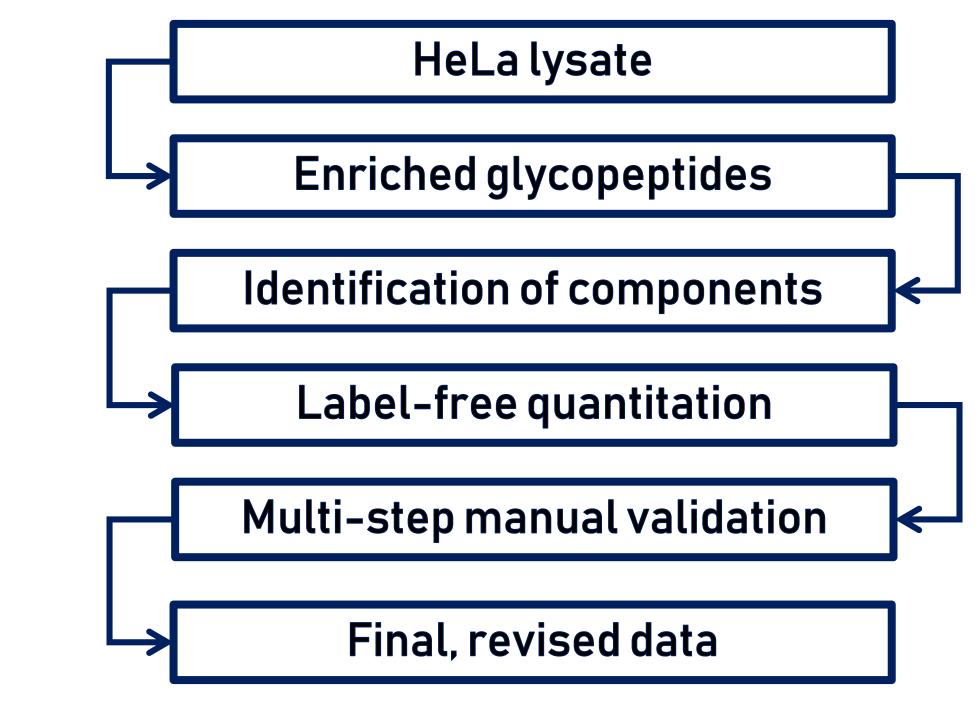
• Data suggested that previous studies on HeLa may have been biased especially released glycan analyes, where the origin of the glycan structures can not be determined

Effects and degree of contamination:

- Approximately the same amount of bovine and human glycopeptides identified in the sample
- The glycosylation pattern of the bovine proteins is very different from the human proteins

Partition of major glycan types by organisms:





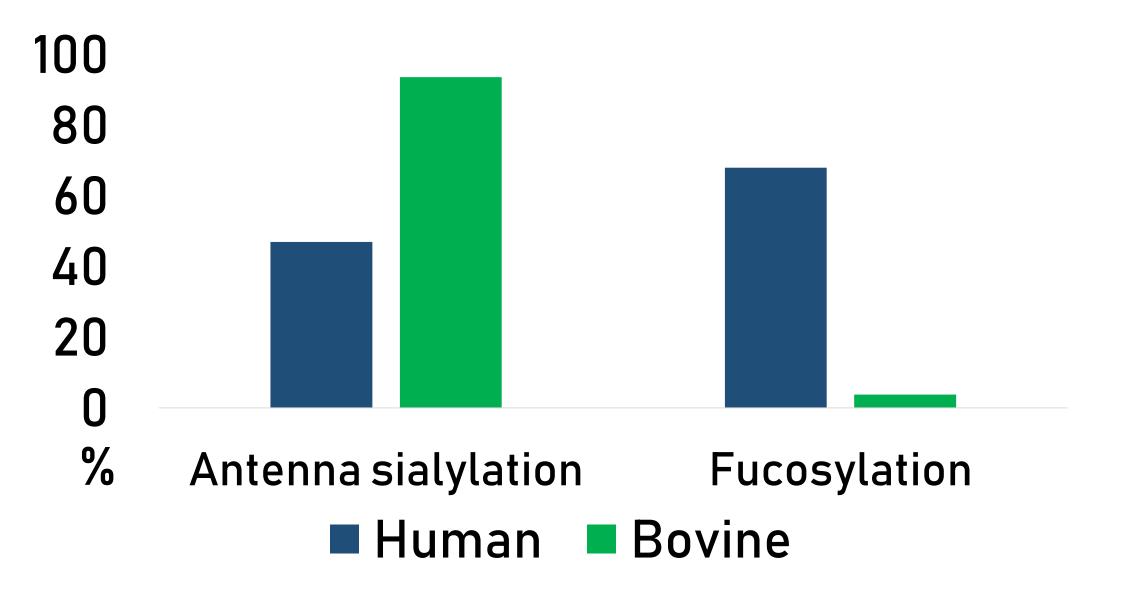
HeLa glycosylation:

 Over 40 glycoproteins identified with 69 glycosylation sites, 28 different glycan structures overall 178 different glycopeptides with high confidence

The identified glycosylation sites:

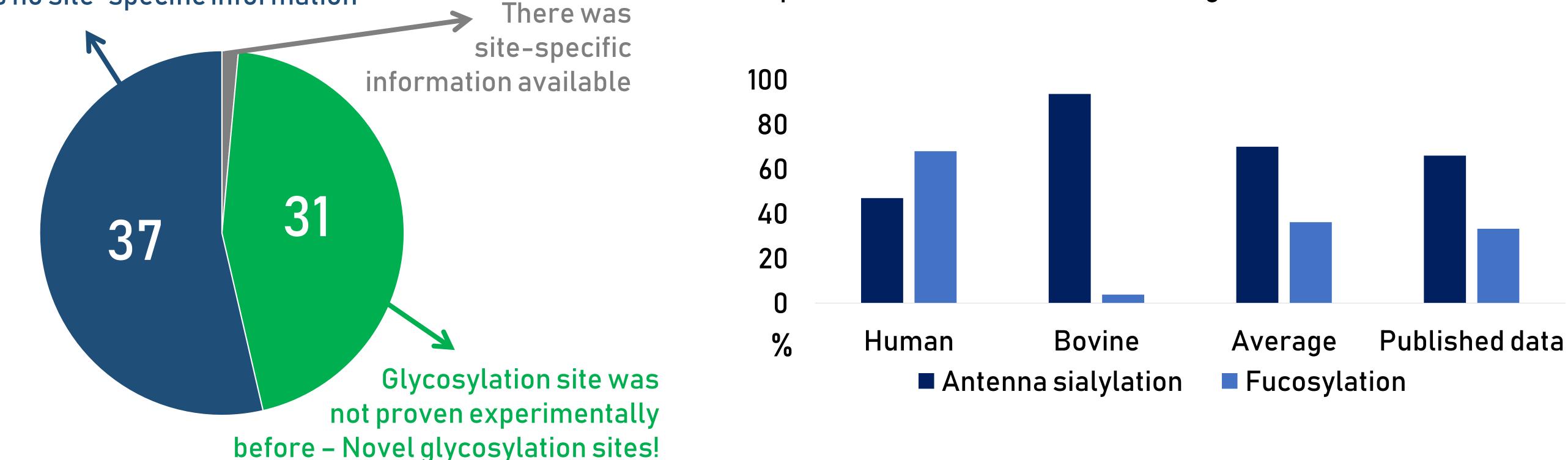
Glycosylation site was known before but there was no site-specific information

Sialylation and fucosylation by organisms:



Comparison of results with published data:

- Comparison of our experimental data on the N-glycosylation pattern of HeLa proteins with a previously published released glycan analysis of HeLa* indicate significant bias
- Results clearly show, that due to the high amount of bovine protein contaminants the sialylation and fucosylation levels published are similar to our averaged human & bovine data



REFERENCES: *: Gao, Wenjie, et al. "A facile method for cellular N-glycomic profiling by matrix-assisted laser desorption/ionization mass spectrometry." RSC Advances 7.57 (2017): 35687-35693. ACKNOWLEDGEMENTS: LT and KV are grateful for funding from the National Research Development and Innovation Office (NKFIH PD-121187 and NKFIH K-119459). LT is grateful for support from the János Bolyai Research Scholarship of the Hungarian Academy of Sciences. SS is grateful for the organizers of MSACL Salzburg for the MSACL Young Investigator Educational Grant which made this presentation possible.